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# **Development of Cell-based Assays for the Identification and Characterization of** Inhibitors of Mutant G323E Hypoxia Inducible Factor- $2\alpha$ (HIF- $2\alpha$ ).



Cesar Meleza, Lunda Shen, Ritu Kushwaha, Amber Pham, Yasmin Esparza, Steven Shia, Mariya Morar, Adam Park, Kenneth V Lawson, Kelsey E Sivick Gauthier, Hyock J Kwon, Xiaoning Zhao Arcus Biosciences, Inc., 3928 Point Eden Way, Hayward, CA 94545, USA

# INTRODUCTION

The role of Hypoxia Inducible Factor- $2\alpha$  (HIF- $2\alpha$ ) in the hypoxia response pathway has garnered attention as a target for the treatment of von Hippel-Lindau (VHL) mutated clear cell renal cell carcinoma (ccRCC). Belzutifan is a HIF-2α small molecule inhibitor, approved by the FDA in 2021, for the treatment of VHL disease. Reported objective responses to HIF-2 $\alpha$  inhibitors tend to be long and potential mechanisms of adaptive resistance have not been reported; however, during the development of first-generation HIF-2α inhibitor, PT2385, drug resistance was observed in a patient that led to the identification of the G323E HIF-2 $\alpha$ gatekeeper mutant.<sup>1</sup> We developed cell-based assays to determine the feasibility of generating compounds that inhibit HIF-2α with the G323E mutation. Using previously reported normoxia stable HIF-2α mutations, HEK-293 cells were engineered to express stable wild type or G323E mutant HIF-2α. These cells were transfected with a hypoxia response element (HRE) luciferase reporter to measure the ability of compounds to inhibit the HIF-2 $\alpha$  luciferase response. Identified inhibitors were further characterized by VEGF secretion in wild type or CRISPR generated 786-O G323E mutant renal cells and in a protein thermal shift assay (TSA). The assays developed provide a platform for screening against and identifying small molecule inhibitors of the G323E HIF-2α mutant.





# RESULTS

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# HEK-293 Luciferase Reporter

- Figure 4a) and 4b) Select compound dose responses in reporter assays.
- Compound Z demonstrates inhibition against the stabilized HIF-2 $\alpha$  and the stabilized G323E HIF-2α.
- Compound X is selective for wild type HIF-2a and not G323E.

Potency correlation of Figure 4c) select inhibitors against the stable wild type and stable G323E HIF-2 $\alpha$ .

 Compounds demonstrate selectivity towards the wild type HIF- $2\alpha$ .



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# • Wide range of potencies is observed against the G323E mutant.





# **MATERIALS AND METHODS**

# HEK-293 HIF-2α Luciferase Reporter Assay

- Figure 1a) The P405A/P531A mutations have been reported to stabilize HIF-2α in normoxia.<sup>2</sup>
- Constructs for stable or G323E HIF-2α were generated with a Cterminal HiBit tag to allow monitoring of protein expression and clone selection.
- HEK-293 cells were transfected with either stable or G323E HIF- $2\alpha$  to achieve protein stability under normoxia.
- Clones expressing stabilized HIF-2 $\alpha$  with or without G323E mutation were identified utilizing the Nano-Glo® HiBiT Lytic Detection System (Promega).

### Figure 1b) Stabilized positive expressing clones were transfected with HRE luciferase reporter to measure HIF-2 $\alpha$ activity.

• Endogenous HIF-2α from HEK-293 is quickly degraded in normoxia condition. Minimal HRE-reporter activity is detected in

# 786-O VEGF Secretion

Figure 5b) Select **5**a) and compound dose response in VEGF secretion assay.

• The same compounds dosed in the reporter assay recapitulate their results in the 786-O VEGF assay.

#### Figure 5c) Compound correlation between wildtype assays.

- Compounds appear slightly more potent in the VEGF assay.
- Good correlation is observed between select inhibitors comparing the reporter assay to the VEGF assay.

# Figure 5d) Compound correlation between G323E assays

• Good correlation is observed for select compounds when comparing the luciferase reporter. assay to the 786-O VEGF.

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### 786-O VEGF Secretion Assay

Figure 2a) 786-O renal cells lack a functional VHL gene allowing for constitutive HIF-2 $\alpha$  activity. This endogenous HIF-2 $\alpha$  activity made for an ideal system to measure HIF-2 $\alpha$  inhibition via downstream VEGF secretion.

• Utilizing CRISPR, a 786-0 cell line was generated with the G323E HIF-2α mutation.

# Figure 2b) VEGF secretion measured by AlphaLISA (Perkin Elmer).

• Wild type and CRISPR generated G323E 786-O cells were treated with HIF-2α inhibitors over 48 hours. After compound treatment, the supernatant was collected, and VEGF secretion was measured utilizing the VEGF AlphaLISA.



# HIF-2α Thermal Shift Assay

Figure 3) Using purified PAS-B domain for wild type or G323E HIF-2α inhibitor binding was measured using a thermal shift assay.

- In the presence of Protein Thermal Shift (Thermo) dye, protein is exposed to an increasing temperature gradient. As protein denatures, the dye binds to exposed hydrophobic regions increasing fluorescence generating a melting curve.
- The presence of inhibitor binding stabilizes the protein at higher temperatures, shifting the melting point (Tm) for the protein. This shift in Tm is proportional to

parental cells.

- Cells were incubated with compound for 24 hours.
- After 24 hours, One-Glo (Promega) was added, and luminescence read



#### 786-O Wt VEGF Ξ 786-0 G323E VEGF Average $IC_{50}$ (M) Average $IC_{50}$ (M)





# **Thermal Shift Assay**

Figure 6a) and 6b) Thermal shift assay melting curve of PAS-B wildtype or G323E domain in the presences of select inhibitors.

- Compound X shows minimal shift in  $\Delta$ Tm against the G323E mutant compared to wild type HIF-2 $\alpha$ .
- Compound Z generates a significant ΔTm against both the wild type and the G323E mutant confirming binding of inhibitor to the PAS-B domain of both wild type and G323E HIF-2 $\alpha$ .

# CONCLUSION

- Expression of normoxia stable HIF-2α in HEK-293 cells and subsequent development of luciferase reporter assay allowed for testing and identification of inhibitors against both wild type and G323E HIF-2 $\alpha$ .
- In addition, inhibition of endogenous HIF-2α activity was measured in CRISPR generated 786-O using VEGF AlphaLISA.
- The data recapitulates the results of both the wildtype and G323E across cell-based assays.
- Binding of inhibitors to G323E mutants was confirmed by thermal shift assay and subsequently compound binding mode was identified via X-ray crystallography.<sup>3</sup>

#### References

- Courtney K. et al. Clin Cancer Res 15 February 2020; 26 (4): 793–803.
- 2) Fujita, N. et al. J Bone Miner Res. 2012 Feb; 27(2): 401–412.





3) Shia, S et al. Partially open conformation of the G323E mutated HIF-2α PASB domain captured by X-ray crystallography. In: Proceedings of the AACR-NCI-EORTC Virtual International Conference on Molecular Targets and Cancer Therapeutics; 2023 Oct 11-15; Boston, MA. Philadelphia (PA): AACR; Mol Cancer Ther 2023;22.

4) Illustrations created with BioRender.com (2024).