

# Small-Molecule Inhibitors of TNF Reduce Pro-inflammatory Disease Activity while Preserving TNFR2 Signaling, Which Maintains Regulatory T Cell Activity and Prevents Aberrant T Cell Cytokine Production



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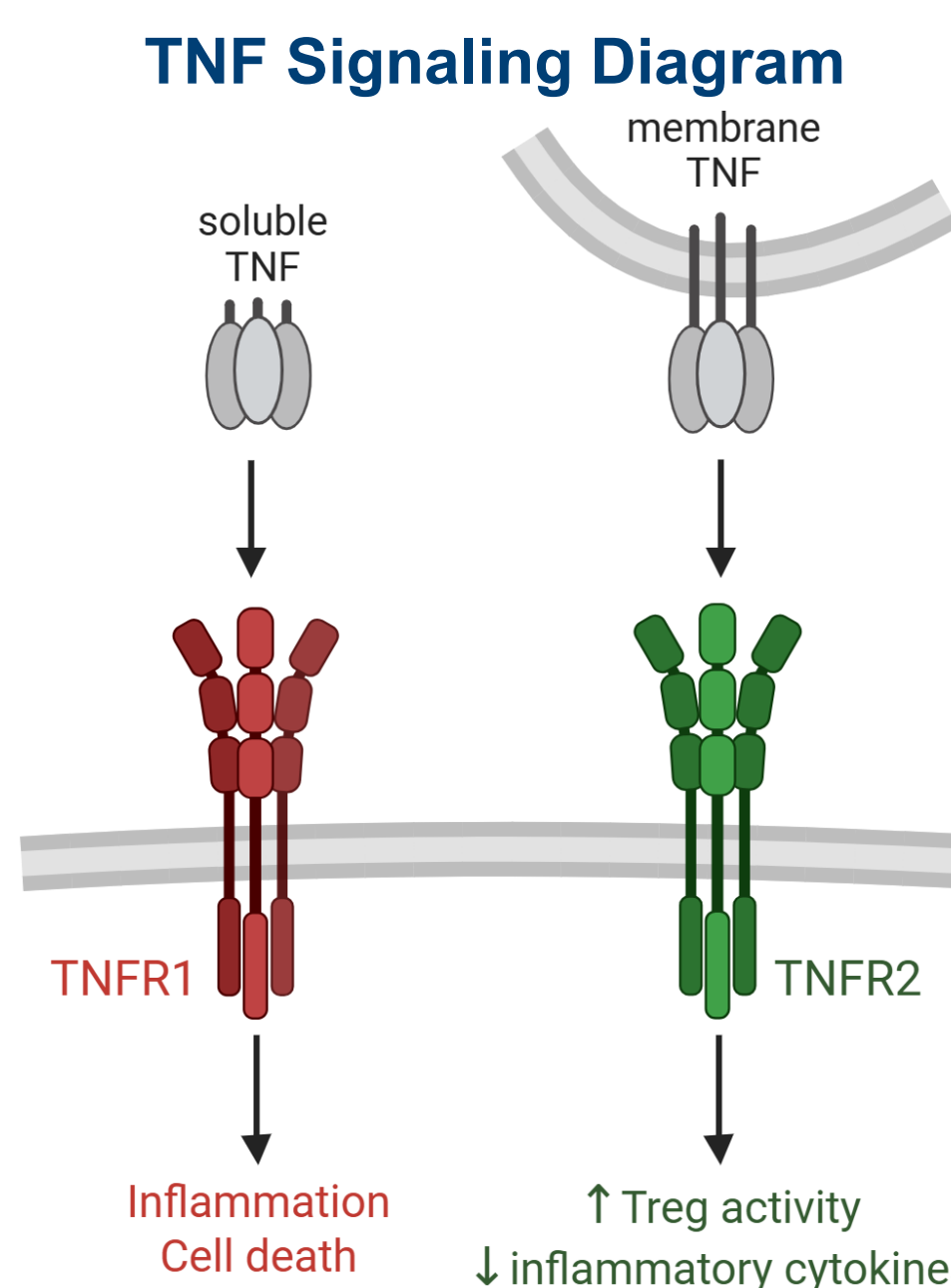
## Background

Tumor necrosis factor (TNF) plays a critical role in inflammation and anti-TNF biologics are effective in treating inflammatory diseases such as rheumatoid arthritis, psoriasis, and inflammatory bowel disease.

TNF is initially expressed as a membrane-bound precursor that is subsequently cleaved by the metalloprotease TACE (ADAM17) to release its soluble form.

Soluble TNF (sTNF) preferentially activates pro-inflammatory TNFR1 signaling while TNFR2 requires membrane TNF (mTNF) for efficient downstream signaling.

Herein, we demonstrate that small-molecule TNF inhibitors block sTNF/TNFR1 signaling while preserving beneficial mTNF/TNFR2 signaling that is blocked by conventional biologic antagonists.



**Figure 1.** Soluble TNF activates TNFR1 which mediates inflammation and cell death. Membrane-bound TNF activates TNFR2 to promote Treg activity and prevent pathogenic cytokine production.

## Methods

HEK293 cells stably expressing an NF- $\kappa$ B-SEAP reporter or K562 cells transiently transfected with an NF- $\kappa$ B luciferase reporter plasmid were used to evaluate TNF receptor activity. TNFR1<sup>KO</sup> K562 cells were generated using CRISPR.

HeLa cells were engineered to express a non-cleavable form of TNF (mTNF) and cultured with reporter cells or isolated human regulatory T cells (Treg) for 4-36 hours.

Primary human vascular endothelial cells (HUVEC) were cultured in presence of small-molecule inhibitors of TNF or adalimumab in presence of 0.125 ng/ml TNF for 24 hours. Cell supernatants were quantified for IL-8 using Meso Scale Discovery (MSD) assays.

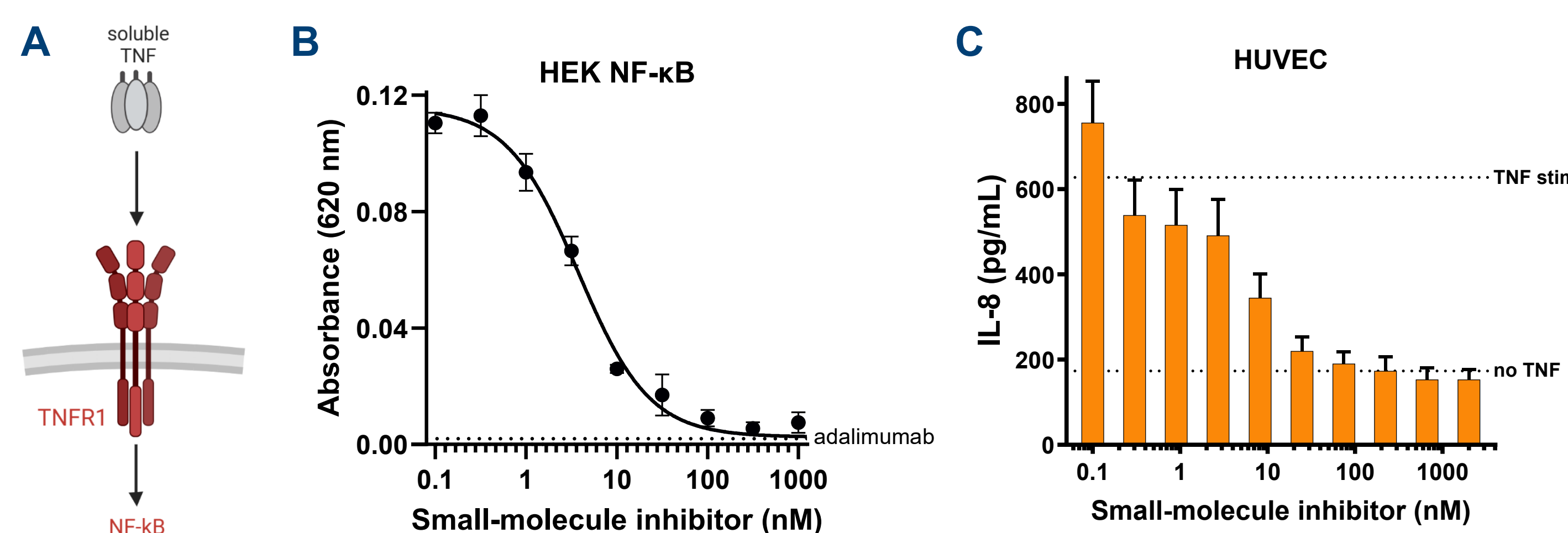
Tregs were isolated using the CD4<sup>+</sup>CD127<sup>low</sup>CD25<sup>+</sup> Treg isolation kit from Stem Cell Technologies and cultured with mTNF-expressing cells for 36 hours. Cells were collected for flow cytometry, and cytokines were measured in cell supernatants by cytokine bead array.

For murine arthritis, anti-collagen antibodies were purchased from Chondrex or chicken collagen II in complete or incomplete Freund's adjuvant was purchased from Hooke laboratories.

Cell culture experiments were performed with example 6 from Pat. Appl. WO 2018/197503 A1. Murine experiments were performed with an in-house synthesized small-molecule inhibitor.

## Results

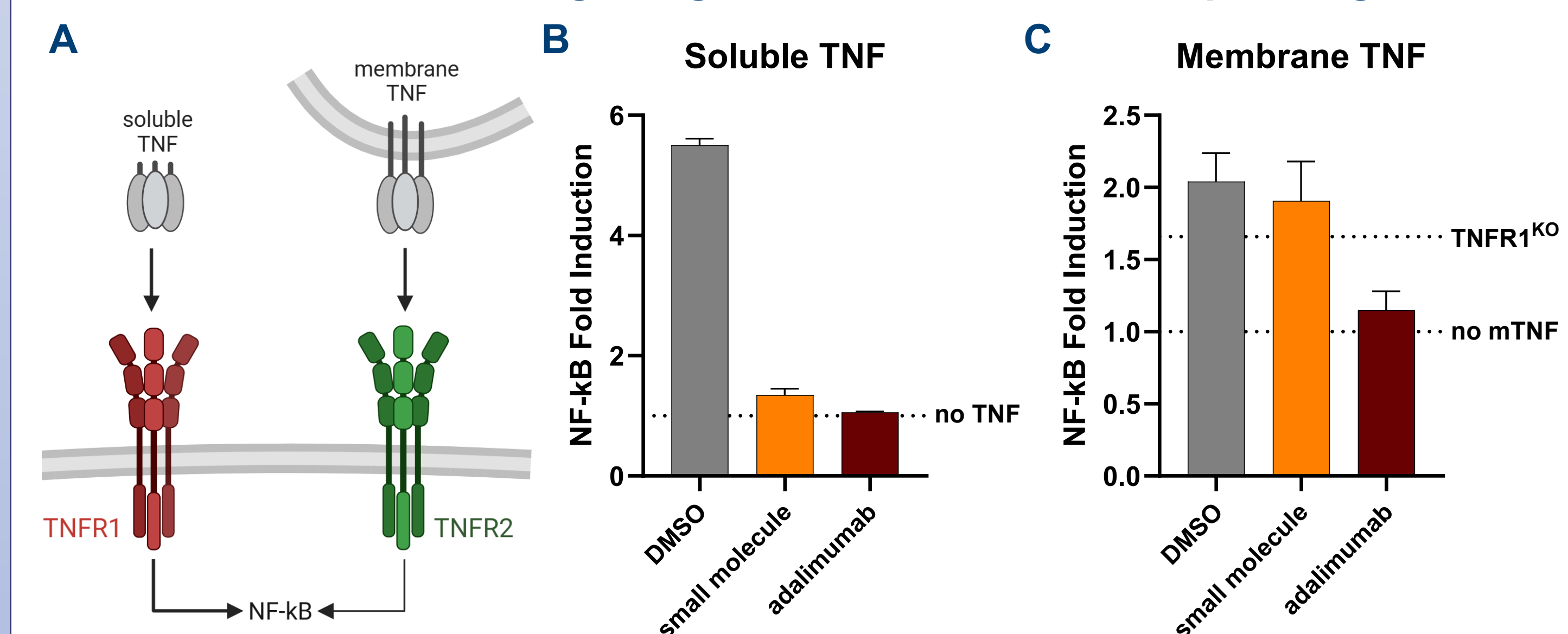
### Small-Molecule TNF Inhibition Fully Blocks Soluble TNF-Driven Activity



**Figure 2.** (A) Model of soluble TNF signaling through TNFR1 to activate NF- $\kappa$ B. (B) NF- $\kappa$ B reporter HEK293 cells were incubated with 0.125 ng/mL of TNF and increasing concentrations of small-molecule inhibitor, reporter activity was measured by luminescence. (C) Secreted IL-8 levels were measured in HUVEC media at 24 hours post incubation with soluble TNF and increasing concentrations of small-molecule inhibitor. N=4 donors

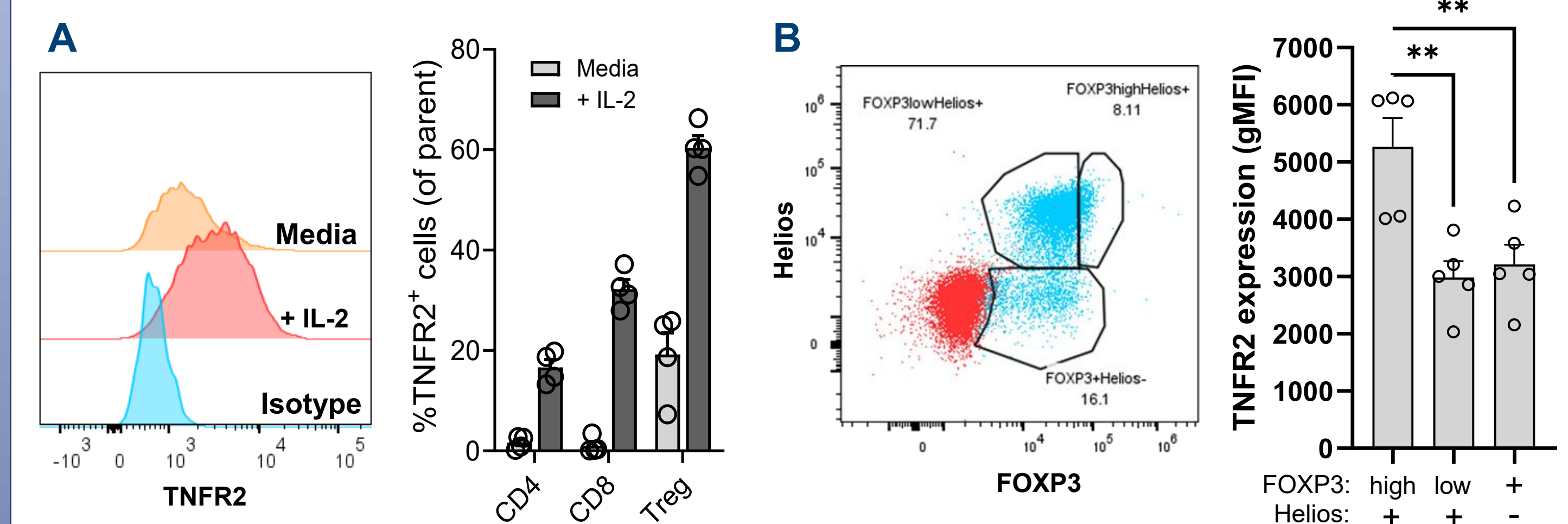
## Results

### Small-Molecule TNF Inhibition Fully Blocks Soluble TNF Signaling But Does Not Block Membrane TNF Signaling in TNFR1/TNFR2 Dual-Expressing Cells



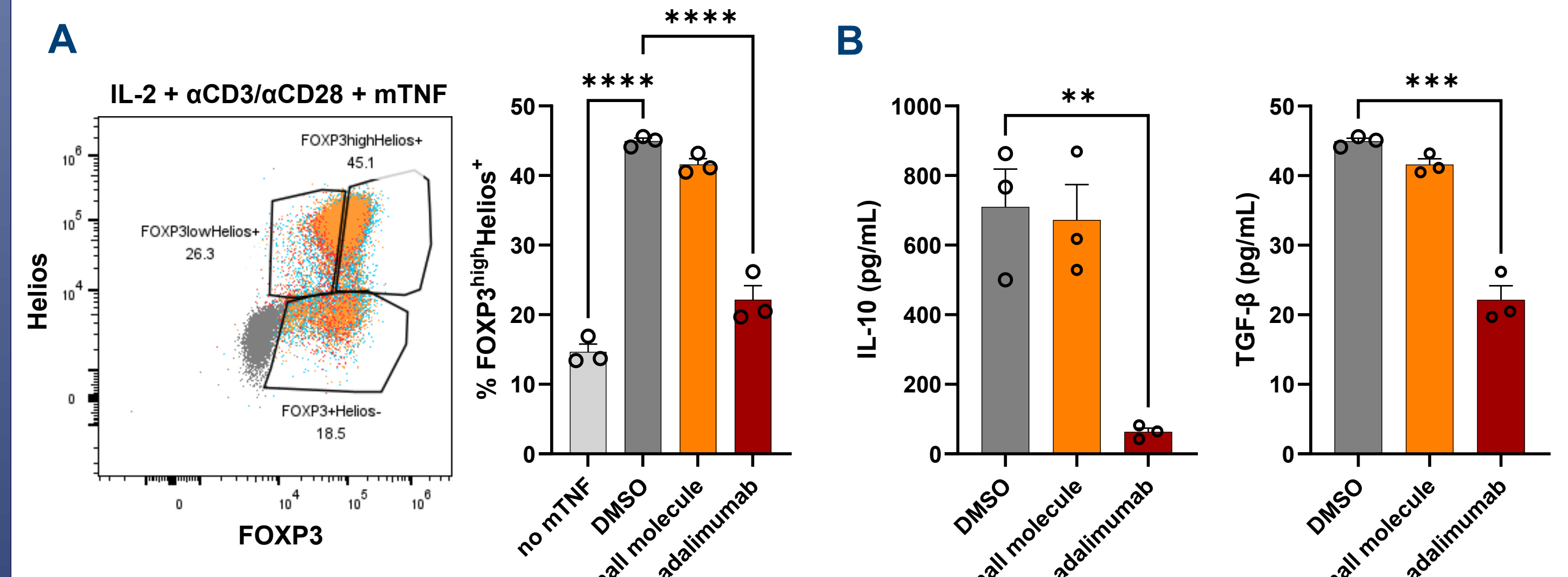
**Figure 3.** (A) Model of NF- $\kappa$ B activation by soluble or membrane TNF. (B) Both a small-molecule TNF inhibitor and adalimumab inhibit soluble TNF activation of NF- $\kappa$ B in HEK293 cells. (C) Adalimumab, but not the small-molecule TNF inhibitor, suppressed membrane TNF-stimulated NF- $\kappa$ B activation through TNFR2 in K562 cells. TNFR1 was blocked using an anti-TNFR1 antibody.

### IL-2 Signaling Drives Expression of TNFR2 on Human T Helper Cells and is Abundantly Expressed on Highly Suppressive Regulatory T Cells



**Figure 4.** (A) Total T cells were isolated from PBMC and cultured for 48 hours with or without IL-2 prior to surface TNFR2 analysis by flow cytometry. A greater percentage of T regulatory cells express surface TNFR2 compared to CD4<sup>+</sup> and CD8<sup>+</sup> T cells. (B) TNFR2 expression was evaluated on freshly isolated healthy human Tregs. The highly suppressive Foxp3<sup>high</sup>Helios<sup>+</sup> Tregs showed the most abundant TNFR2 expression. N=4 human donors, \*\*p<0.01

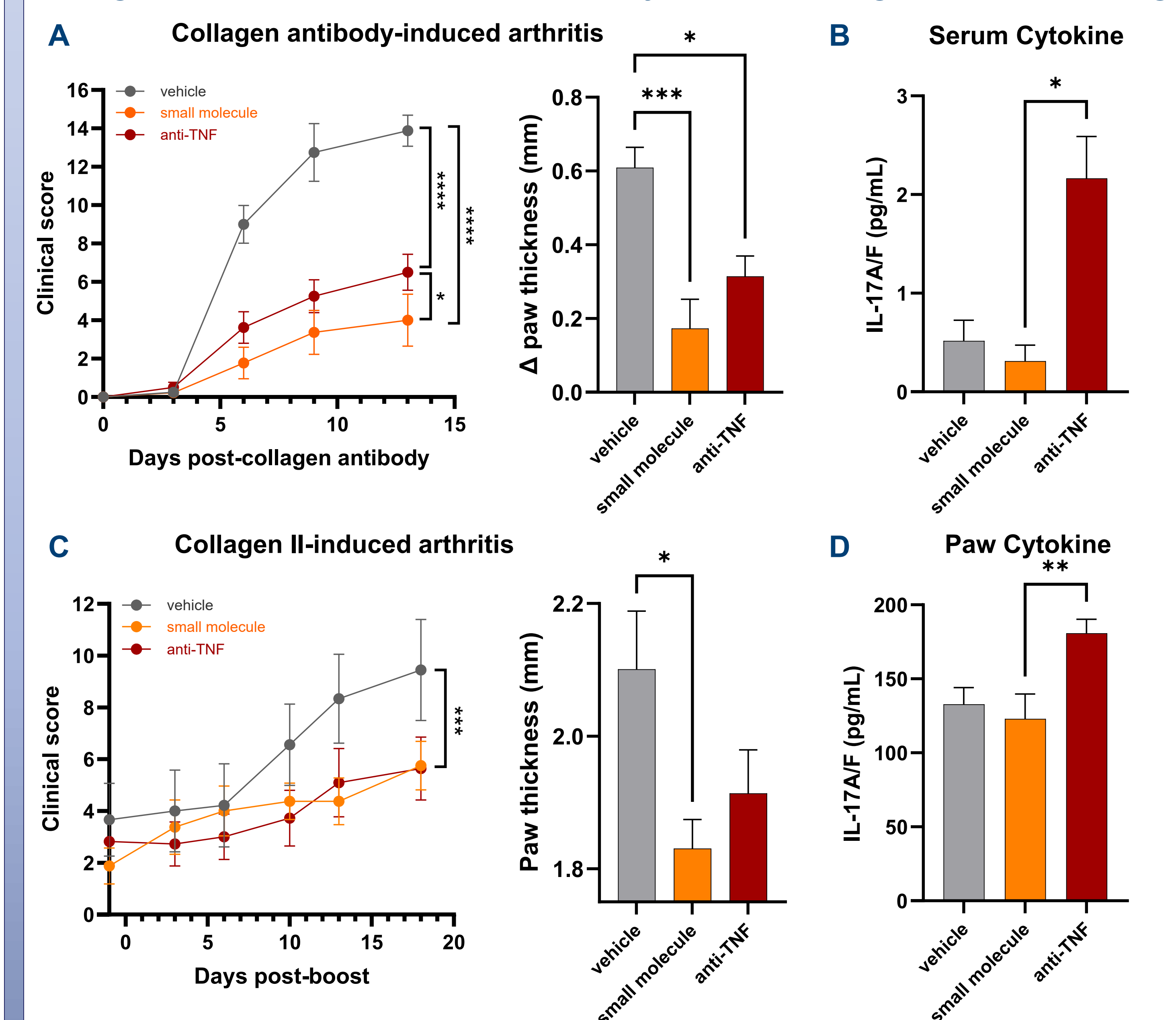
### Membrane TNF-Signaling through TNFR2 Enhances Treg Activity Which is Blocked by an Anti-TNF Biologic but Not a Small-Molecule Inhibitor



**Figure 5.** (A) Purified human Tregs were stimulated with IL-2 and anti-CD3/anti-CD28 and co-cultured with mTNF-expressing cells in the presence of a small-molecule TNF inhibitor or anti-TNF biologic. (B) Secretion of IL-10 and TGF- $\beta$  was measured in supernatants by MSD. N=3 healthy human donors, \*\*p<0.01; \*\*\*p<0.001; \*\*\*\*p<0.0001

## Results

### Small-Molecule TNF Inhibition Provides Equivalent Disease Control as a Biologic in Murine Arthritis Models and May Prevent Pathogenic T Cell Skewing



**Figure 6.** (A) Arthritis clinical score and paw thickness were assessed in the collagen antibody-induced arthritis (CAIA) model demonstrating that small-molecule TNF inhibition provides equivalent disease inhibition compared to an anti-TNF biologic. (B) Serum IL-17A/F production is not significantly changed by small-molecule inhibitor but is increased in the anti-TNF biologic group. (C) Clinical score and paw thickness were assessed in the collagen II-induced arthritis (CIA) model. (D) Paw tissue homogenates demonstrated that an anti-TNF biologic increased IL-17A/F production compared to small-molecule inhibitor and the vehicle control. \*p<0.05; \*\*\*p<0.001; \*\*\*\*p<0.0001

## Conclusions

Small-molecule inhibitors of TNF show equivalent blockade of pro-inflammatory sTNF/TNFR1 signaling as anti-TNF biologics but do not block membrane TNF signaling through TNFR2.

Membrane TNF signaling enhances the suppressive capacity of regulatory T cells which is not blocked with a small-molecule TNF inhibitor but is abrogated by an anti-TNF biologic.

In a murine arthritis model, small-molecule TNF inhibition reduces disease activity as well as an anti-TNF biologic and reduces inflammatory cytokine production, potentially through TNFR2 signaling.

**Figure 7.** Anti-TNF biologics block both soluble and membrane TNF signaling whereas small-molecule inhibitors block only soluble TNF, leaving TNFR2 signaling available to promote Treg activity and suppress inflammatory cytokine production

